

RESEARCH

Open Access

# Evolving thermostability in mutant libraries of ligninolytic oxidoreductases expressed in yeast

Eva García-Ruiz<sup>1,2</sup>, Diana Maté<sup>1</sup>, Antonio Ballesteros<sup>1</sup>, Angel T Martínez<sup>2</sup>, Miguel Alcalde<sup>1\*</sup>

## Abstract

**Background:** In the picture of a laboratory evolution experiment, to improve the thermostability whilst maintaining the activity requires of suitable procedures to generate diversity in combination with robust high-throughput protocols. The current work describes how to achieve this goal by engineering ligninolytic oxidoreductases (a high-redox potential laccase -HRPL- and a versatile peroxidase, -VP-) functionally expressed in *Saccharomyces cerevisiae*.

**Results:** Taking advantage of the eukaryotic machinery, complex mutant libraries were constructed by different *in vivo* recombination approaches and explored for improved stabilities and activities. A reliable high-throughput assay based on the analysis of  $T_{50}$  was employed for discovering thermostable oxidases from mutant libraries in yeast. Both VP and HRPL libraries contained variants with shifts in the  $T_{50}$  values. Stabilizing mutations were found at the surface of the protein establishing new interactions with the surrounding residues.

**Conclusions:** The existing tradeoff between activity and stability determined from many point mutations discovered by directed evolution and other protein engineering means can be circumvented combining different tools of *in vitro* evolution.

## Background

During the last couple of decades, thermostability has been considered by many as a key feature in terms of protein robustness, evolvability and catalytic function [1-4]. From a practical point of view, the engineering of thermo-tolerant biocatalysts is highly desirable since transformations at high temperatures intrinsically supply a box-set of key biotechnological advantages (higher entropies -better reaction yields-, solubilisation of hydrophobic compounds or low levels of microbial side-contamination, among others). Besides, thermostable enzymes are typically tolerant to many other harsh conditions often required in industry, such as the presence of organic co-solvents, extreme pHs, high salt concentrations, high pressures, etc [5,6]. Few exceptions aside [7,8], the discovery of stabilizing mutations is not always straightforwardly accomplished without significant drops in turnover rates [9]. Most of these mutations, which establish new interactions by salt bridges, hydrogen bonds, hydrophobic contacts or even disulfide bridges, are placed either at the protein surface or in internal

cores pursuing the tightly packing of the tertiary protein structure in order to prevent unfolding and denaturation under extreme environments [10]. On the contrary, improvements in activity are generally accomplished by introducing beneficial but destabilizing mutations in *hot* regions for catalysis (substrate binding sites, channels of access to the active pockets) although sometimes distant mutations can also vary the catalytic function by altering the dynamics and geometry in the protein scaffold [3]. There are several examples in literature about the stabilization of enzymes by directed evolution or rational design but unfortunately, main constraints still remain from the lack of appropriate methods to recreate diversity in conjunction with reliable screening strategies, especially if one wants to surpass the existing tradeoff between activity and thermostability for many single residue substitutions [10-20].

Among the enzymes forming the ligninolytic system of white-rot fungi (*i.e.* involved in lignin biodegradation), high redox potential laccases HRPL (EC 1.10.3.2) and peroxidases, including versatile peroxidases (VP; EC 1.11.1.14) are outstanding biocatalysts finding potential applications in paper pulp bleaching and functionalization,

\* Correspondence: malcalde@icp.csic.es

<sup>1</sup>Department of Biocatalysis, Institute of Catalysis, CSIC, 28049 Madrid, Spain

bioremediation, organic synthesis, food and textile industries, nanobiodevice construction and more [21-23]. Indeed, HRPL can oxidize dozens of different compounds releasing water as the only by-product and in the presence of redox mediators (diffusible electron carriers from natural or synthetic sources) their substrates specificities are further expanded [24,25]. On the other hand, VP (with redox potential above +1000 mV) shares the catalytic features of lignin and manganese peroxidase in terms of substrate specificity, together with the ability to oxidize phenols and dyes characteristic of low redox-potential peroxidases. Indeed, the presence of different catalytic sites in a small and compact protein structure (around 300 amino acids) makes VP an ideal platform for laboratory evolution strategies [23,26,27].

Here, we have employed these two enzymatic systems as departure points to improve their protein thermostability by directed evolution. VP and HRPL were functionally expressed in yeast and mutant libraries were constructed combining several methodologies of *in vitro* evolution to guarantee the library complexity, favoring the selection of optimal crossover events or the discovery of beneficial mutations. Highly functional/soluble expressed mutants were stressed under high temperatures and explored for activity and stability. The analysis of the data from screening (ratio residual activity/initial activity in combination with the  $T_{50}$  values) enabled us to discover stabilizing mutations in both systems.

## Materials and methods

HRPL from basidiomycete PM1 [28] (PM1-7H2 mutant) and VP from *Pleurotus eryngii* (10C3, 6B1, 13E4, 6E7 and 11F3 mutants of the allelic variant VPL2, GenBank AF007222) were used as parent types for library construction. Both systems are from previous engineering work by several rounds of directed evolution in *S. cerevisiae* including the replacement of their original native signal sequences by the alpha factor prepro-leader, ([29] and unpublished material). ABTS (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)), bovine haemoglobine, Taq polymerase and the *S. cerevisiae* transformation kit were purchased from Sigma-Aldrich (Madrid, Spain). The *E. coli* XL2-blue competent cells and the Genemorph Random mutagenesis kit were from Stratagene (La Jolla, CA, USA). The protease deficient *S. cerevisiae* strain BJ5465 was from LGCPromochem (Barcelona, Spain). The uracil independent and ampicillin resistance shuttle vector pJRoC30 was obtained from the California Institute of Technology (CALTECH, USA), while the zymoprep yeast plasmid miniprep kit, zymoclean gel DNA recovery kit, and the DNA clean and concentrator TM-5 kit were all from Zymo Research (Orange, CA). NucleoSpin Plasmid kit was purchased from Macherey-Nagel (Germany) and the restriction enzymes *Bam*HI and *Xho*I were from New

England Biolabs (Hertfordshire, UK). All chemicals were of reagent-grade purity.

## 1. Culture media

Minimal medium contained 100 mL 6.7% sterile yeast nitrogen base, 100 mL 19.2 g/L sterile yeast synthetic drop-out medium supplement without uracil, 100 mL sterile 20% raffinose, 700 mL *sdd*H<sub>2</sub>O and 1 mL 25 g/L chloramphenicol. YP medium contained 10 g yeast extract, 20 g peptone and *dd*H<sub>2</sub>O to 650 mL. Expression medium contained 720 mL YP, 67 ml 1 M KH<sub>2</sub>PO<sub>4</sub> pH 6.0 buffer, 111 mL 20% galactose, 1 ml 25 g/L chloramphenicol and *dd*H<sub>2</sub>O to 1000 mL. For HRPL the expression medium was supplemented with 2 mM CuSO<sub>4</sub> and 25 g/L ethanol. For VP the expression medium was supplemented with 100 mg/L bovine haemoglobine. YPD solution contained 10 g yeast extract, 20 g peptone, 100 mL 20% sterile glucose, 1 ml 25 g/L chloramphenicol and *dd*H<sub>2</sub>O to 1000 mL. SC drop-out plates contained 100 mL 6.7% sterile yeast nitrogen base, 100 mL 19.2 g/L sterile yeast synthetic drop-out medium supplement without uracil, 20 g bacto agar, 100 mL 20% sterile glucose, 1 mL 25 g/L chloramphenicol and *dd*H<sub>2</sub>O to 1000 ml.

## 2. Library construction for laboratory evolution

### General aspects

Unless otherwise specified, PCR fragments were cleaned, concentrated and loaded onto a low melting point preparative agarose gel and purified using the Zymoclean gel DNA recovery kit (Zymo Research). PCR products were cloned under the control of the Gal 10 promoter of the expression shuttle vector pJRoC30, replacing the corresponding parental gene in pJRoC30. To remove the parental gene, the pJRoC30 plasmid was linearized (with *Xho*I and *Bam*HI for HRPL- and VP-libraries). Linearized vector was concentrated and purified as described above for the PCR fragments.

Mutagenic *StEP* (*Staggered Extension Process*) followed by *in vivo* DNA shuffling and *IvAM* (*In vivo* Assembly of Mutant libraries with different mutational spectra) were used to create the VP and HRPL libraries, respectively, as described below.

### VP Library: mutagenic *StEP* + *in vivo* DNA shuffling

10C3, 6B1, 13E4, 6E7 and 11F3 VP-mutants were used as parental types. *StEP* was performed as reported elsewhere [30] with some modifications. In order to favor random mutagenesis during *StEP*, Taq DNA-polymerase was employed for the PCR reaction along with low concentration of templates to promote the introduction of point mutations during the amplification. The primers used were: RMLN-sense (5'-CCTCTAATACTTTAACGTCAAGG-3') and RMLC-antisense (5'-GGGAGGGC-GTGAATGTAAGC-3'). For the *in vivo* ligation, overhangs of 40 bp and 66 bp that were homologous to

linearized vector were designed. PCR reactions were performed in a final volume of 50  $\mu$ L containing 90 nM RMLN, 90 nM RMLC, 0.3 mM dNTPs, 3% dimethylsulfoxide (DMSO), 0.05 U/ $\mu$ L of Taq polymerase (Sigma), 1.5 mM MgCl<sub>2</sub> and 0.1 ng/ $\mu$ L of 10C3, 6B1, 13E4, 6E7 and 11F3 DNA-template mixture. *StEP* was carried out using a gradient thermocycler (Mycycler, Biorad, USA). The thermal cycling parameters were as follows: 95°C for 5 min (1 cycle), 94°C for 30 s and 55°C for 20 s (90 cycles). Purified PCR products were further recombined by *in vivo* DNA-shuffling [31]. PCR mutated/recombined products were mixed equimolarly (160 ng of each product) and transformed along with linearized vector (ratio PCR product:vector, 4:1) into competent cells using the yeast transformation kit (Sigma). A mutant library of ~2000 clones was explored.

#### **HRPL Library: *IvAM***

*IvAM* (~1300 clones) was performed as reported elsewhere [32] with some modifications. HRPL PM1-7H2 mutant was used as parent type. Mutagenic PCR was carried out using the following thermal cycling parameters: 95°C for 2 min (1 cycle), 94°C for 0.45 min, 53°C for 0.45 min, 74°C for 3 min (28 cycles), 74°C for 10 min (1 cycle). For the Taq library the concentrations of each ingredient in 50  $\mu$ L final volume were as follows: 90 nM RMLN; 90 nM RMLC; 0.1 ng/ $\mu$ L HRPL template; 0.3 mM dNTPs (0.075 mM each); 3% DMSO; 1.5 mM MgCl<sub>2</sub>; 0.01 mM MnCl<sub>2</sub> and 0.05 U/ $\mu$ L Taq polymerase. For the Mutazyme library the concentrations of each reagent in 50  $\mu$ L final volume were as follows: 370 nM RMLN; 370 nM RMLC, 40 ng/ $\mu$ L HRPL template; 0.8 mM dNTPs; 3% DMSO; and 0.05 U/ $\mu$ L Mutazyme DNA polymerase. Taq/MnCl<sub>2</sub> and Mutazyme libraries were equimolarly mixed and transformed along with linearized vector (ratio equimolar library:vector, 8:1) into competent *S. cerevisiae* cells as described above.

### **3. High-throughput thermostability assay**

Individual clones were picked and cultured in 96-well plates (Sero-well, Staffordshire, UK) containing 50  $\mu$ L of minimal medium per well. In each plate, column number 6 was inoculated with standard (parental HRPL or VP), and one well (H1-control) was either not inoculated for HRPL libraries or inoculated with untransformed *S. cerevisiae* cells for VP-libraries. Plates were sealed to prevent evaporation and incubated at 30°C, 225 RPM and 80% relative humidity in a humidity shaker (Minitron-INFORS, Biogen, Spain). After 48 h, 160  $\mu$ L of expression medium were added to each well, and the plates were incubated for 24 h. The plates (master plates) were centrifuged (Eppendorf 5810R centrifuge, Germany) for 5 min at 3000  $\times$  g at 4°C and 20  $\mu$ L of supernatant was transferred from the master plate with the help of a robot (Liquid Handler Quadra 96-320, Tomtec, Hamden, CT,

USA) onto the replica plate. Subsequently, 180  $\mu$ L of stability buffer (10 mM sodium tartrate buffer pH 5.1 for VP-library and 10 mM Britton and Robinson buffer pH 6.0 for HRPL-library) were added to each replica and briefly stirred. Replica plate was duplicated with the help of the robot by transferring 50  $\mu$ L of mixture to a thermocycler plate (Multiply PCR plate without skirt, neutral, Sarstedt, Germany) and 20  $\mu$ L to the initial activity plate. Thermocycler plates were sealed with thermoresistant film (Deltalab, Spain) and incubated at the corresponding temperature using a thermocycler (MyCycler, Biorad, USA). Incubation took place for 10 min (so that the assessed activity was reduced 2/3 of the initial activity). Afterwards, thermocycler plates were placed on ice for 10 min and further incubated for 5 min at room temperature. 20  $\mu$ L of supernatants were transferred from both thermocycler and initial activity plates to new plates to estimate the initial activities and residual activities values by adding ABTS containing specific buffers. For VP-libraries 180  $\mu$ L of 100 mM sodium tartrate buffer pH 3.5 containing 2 mM ABTS and 0.1 mM H<sub>2</sub>O<sub>2</sub> were added to each plate. For HRPL-libraries 180  $\mu$ L of 100 mM sodium acetate buffer pH 5.0 containing 3 mM ABTS were added. Plates were stirred briefly and the absorption at 418 nm ( $\epsilon_{\text{ABTS}^{*+}} = 36,000 \text{ M}^{-1} \text{ cm}^{-1}$ ) was recorded in the plate reader (SPECTRAMax Plus 384, Molecular Devices, Sunnyvale, CA). The plates were incubated at room temperature until a green color developed, and the absorption was measured again. The same experiment was performed for both the initial activity plate and residual activity plate. Relative activities were calculated from the difference between the absorption after incubation and that of the initial measurement normalized against the parental type in the corresponding plate. Thermostability values came from the ratio between residual activities and initial activities values. To rule out false positives, two consecutive rescreenings were carried out according to the protocol previously reported [33] with some modifications. A third rescreening was incorporated to calculate the T<sub>50</sub> of selected mutants.

#### **First rescreening**

Aliquots of 5  $\mu$ L of the best clones were removed from master plates to inoculate 50  $\mu$ L of minimal media in new 96-well plates. Columns 1 and 12 (rows A and H) were not used to prevent the appearance of false positives. After 24 h of incubation at 30°C and 225 RPM, 5  $\mu$ L were transferred to the adjacent wells and further incubated for 24 h. Finally, 160  $\mu$ L of expression medium were added and plates were incubated for 24 h. Accordingly, every single mutant was grown in 4 wells. Parent types were subjected to the same procedure (lane D, wells 7-11). Plates were assessed using the same protocol of the screening described above but including not

only an endpoint assay but also a kinetic assay. In the ABTS kinetic assay, linear absorption increases over a wide range of enzyme concentration (1-20 mU/mL) allowing the estimation of initial rates.

#### **Second rescreening**

an aliquot from the wells with the best clones of first rescreening was inoculated in 3 mL of YPD and incubated at 30°C and 225 RPM for 24 h. Plasmids from these cultures were extracted (Zymoprep yeast plasmid miniprep kit, Zymo Research). As the product of the zymoprep was very impure and the concentration of extracted DNA was very low, the shuttle vectors were transformed into super-competent *E. coli* cells (XL2-Blue, Stratagene) and plated onto LB-amp plates. Single colonies were picked and used to inoculate 5 mL LB-amp media and were grown overnight at 37°C and 225 RPM. Plasmids were then extracted (NucleoSpin® Plasmid kit, Macherey-Nagel, Germany). *S. cerevisiae* was transformed with plasmids from the best mutants and also with parent type. Five colonies of every single mutant were picked and rescreened as described above (using both end-point and kinetic assays).

#### **Third rescreening ( $T_{50}$ determination)**

fresh transformants of selected mutants and parent types were cultivated (10 mL) in 100 mL flask for VP and HRPL production. Supernatants were subjected to a thermostability assay to accurately estimate their  $T_{50}$  using 96/384 well gradient thermocyclers (Mycycler, Biorad, US). Appropriate dilutions of supernatants were prepared with the help of the robot in such a way that aliquots of 20  $\mu$ L give rise to a linear response in kinetic mode. 50  $\mu$ L (from both selected mutants and parent types) were used for each point in the gradient scale. A temperature gradient profile ranging from 30 to 90°C was established. After 10 min of incubation, samples were removed and chilled out on ice for 10 min. After that, samples of 20  $\mu$ L were removed and incubated at room temperature for 5 min. Finally, samples were subjected to the same ABTS-based colorimetric assay described above for the screening. Thermostabilities values were deduced from the ratio between the residual activities incubated at different temperature points and the initial activity at room temperature.

#### **4. Determination of thermostabilities in VP and HRPL parent types**

Thermostabilities of 7H2-HRPL and 10C3-VP mutants were assessed mimicking the growth conditions established for the screening assay as described above. Two 96 well-plates containing 50  $\mu$ L minimal media were inoculated with 7H2 and 10C3 respectively and cultivated until reaching functional expression following the conditions used for the assay. Afterwards, supernatants of 7H2 and 10C3 were pooled and employed to estimate

their respective thermostabilities with the gradient thermocycler. The gradient of temperature was set at the following points (in°C): 30.0, 31.7, 34.8, 39.3, 45.3, 49.9, 53.0, 55.0, 56.8, 59.9, 64.3, 70.3, 75.0, 78.1 and 80 for the VP mutant and 35.0, 36.7, 39.8, 44.2, 50.2, 54.9, 58.0, 60.0, 61.1, 63.0, 65.6, 69.2, 72.1, 73.9, 75.0, 76.2, 78.0, 80.7, 84.3, 87.1, 89.0 and 90.0 for the HRPL mutant. The protocol followed the general rules described for the third re-screening.

#### **5. DNA sequencing**

Plasmid-containing variant HRPL and VP genes were sequenced by using a BigDye Terminator v 3.1 Cycle Sequencing Kit. Primers were designed with Fast-PCR software (University of Helsinki, Finland). Primers used for VP variants were: RMLN; 3R-direct (5'-GTTCCAT-CATCGCGTTTCG-3'); 5F-reverse (5'-GGATTCCTT-TCTTCTTGG-3') and RMLC. For HRPL primers were: RMLN; PM1FS (5'-ACGACTTCCAGGTCCCTGAC-CAAGC-3'); PM1RS (5'-TCAATGTCCGCGTTTCG-CAGGGA-3') and RMLC.

#### **6. Protein modelling**

We carried out a search in the Protein Data Bank for proteins with known structural homology to laccase PM1. The most similar protein to PM1 was a laccase from *Trametes trogii* (crystal structure solved with a resolution of 1.58 Å), showing 97% sequence identity (PDB id: 2hrgA) [34]. A model from the Swiss-Model protein automated modelling server was generated <http://swissmodel.expasy.org/> and analyzed with DeepView/Swiss-Pdb Viewer.

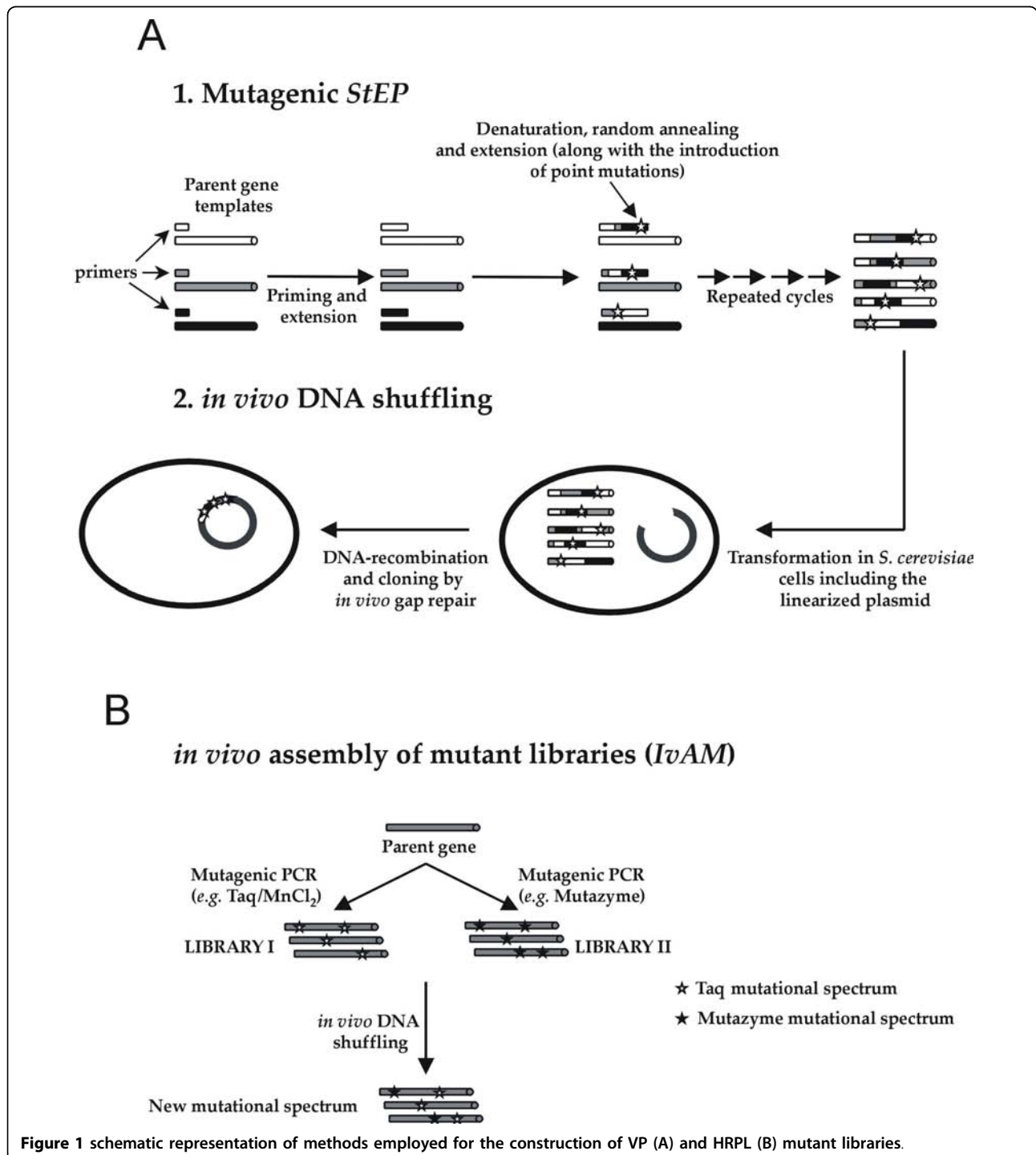
### **Results and discussion**

#### **1. Library construction**

VP and HRPL variants come from laboratory evolution approaches to be functionally expressed in *Saccharomyces cerevisiae* ([29] and unpublished material). Therefore, the array of variants used as starting points for enhancing thermostability, in principle only were going to harbor mutations which conferred higher secretion levels and/or better kinetics. Both VP and HRPL native signal peptides were replaced by the  $\alpha$ -factor pre-proleader to promote secretion in yeast [35]. Thus, the whole fusion proteins ( $\alpha$ -VP and  $\alpha$ -HRPL) were subjected to directed evolution in order to enhance their functional expression; *i.e.* the accumulation of neutral and beneficial mutations took place both in the  $\alpha$ -factor pre-proleader as well as in mature protein. If high fidelity DNA recombination strategies are used, where only accumulated mutations recombine each other based on the number of crossovers generated in the protocol, it is highly likely that the discovery of thermostable variants become a complex task, since all these mutations have been

unmasked from improvements in total activities; *i.e.* the sum of secretion levels plus the specific activity. Hence, we decided to tackle the library design by combining the already accumulated mutations with the generation of new ones randomly incorporated in the frame of mutagenic DNA recombination fashion approaches (Fig. 1). For the VP library the well known *Staggered Extension*

*Process (StEP)* [30] was slightly modified by decreasing templates concentration with the aim of provoking amplification mistakes by the Taq DNA-Polymerase which has a high intrinsic error rate because of the lack of 3'-5' proofreading exonuclease activity. The mutational rate was adjusted to 1-2 mutations per whole gene [36]. Afterwards, and instead of *in vitro* ligating the *StEP*

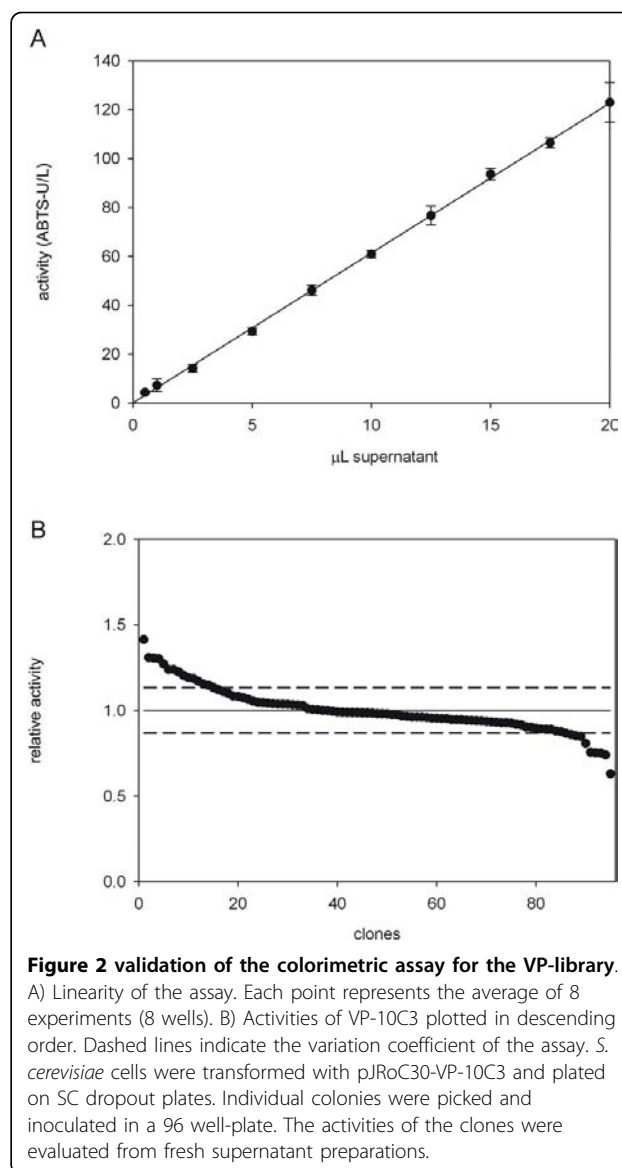


products with the linearized plasmid to give rise to individual autonomously replicating vectors as usually proceeds, we thought that might be interesting to further recombine the pool of mutagenized/crossover containing genes in an *in vivo* approach with the aim of expanding the library diversity. Thus, *StEP* products were further subjected to an *in vivo* DNA-shuffling process [31,37,38] taking advantage of the high level of homologous recombination of *S. cerevisiae* apparatus as described in the Material & Methods section. With this strategy, new crossover events were generated that otherwise hardly could be achieved by using conventional *in vitro* or *in vivo* recombination methods independently. In the case of HRPL library, the only starting template gene was subjected to the *in vivo* assembly of mutant libraries with different mutational spectrum (*IvAM*) [32], to guarantee the enrichment of the library in different mutational types and bias. With this simple approach, the homologous recombination, reparation and *in vivo* cloning of mutant genes along with their *in vivo* DNA-shuffling, in one single step were achieved.

## 2. High-throughput screening assay

ABTS was chosen as substrate for the screening assay since it has a reliable response, high sensitivity and hardly interferes with the main components of culture broth. Indeed, in our former works ABTS was validated in the frame of directed laccase evolution for several purposes [33,38-40]. However, the response of this substrate against the secreted VP mutants by *S. cerevisiae* was unknown and therefore we had to validate it before starting the thermostability studies. First, the relationship between absorbance and VP concentration was evaluated. Fresh transformants containing VP-10C3 mutant (the best parent type) were inoculated in a 96-well plate, VP-10C3 was expressed and different volumes of supernatant were assessed. A linear behavior between the amount of VP and the response of the assay was found (Fig. 2A). The coefficient of variation for the ABTS in kinetic mode was 13%, which is an acceptable value to study mutagenic libraries engineered for directed evolution experiments [9] (Fig. 2B).

For the thermostability assay, we first had to set the temperature under which the mutant libraries would be stressed. Accordingly, the two best parent types of both VP and HRPL libraries were tested in order to know their respective thermostabilities. Taking into account that micro-fermentation conditions (*i.e.* in 96 well plates) were far away from an ideal large-scale fermentation in terms of oxygen availability, surface stirring and length of incubation time, VP-10C3 and HRPL-7H2 mutants were produced under such a limited growth conditions and evaluated to obtain the closest real value of thermostability in the screening assay



**Figure 2 validation of the colorimetric assay for the VP-library.** A) Linearity of the assay. Each point represents the average of 8 experiments (8 wells). B) Activities of VP-10C3 plotted in descending order. Dashed lines indicate the variation coefficient of the assay. *S. cerevisiae* cells were transformed with pJRoC30-VP-10C3 and plated on SC dropout plates. Individual colonies were picked and inoculated in a 96 well-plate. The activities of the clones were evaluated from fresh supernatant preparations.

(further details in Material & Methods section). In principle, an appropriate enzyme heat treatment for the screening is generally chosen so that the residual activity is about one-third of the initial activity [9]. At 60°C and 68°C, VP-10C3 and HRPL-7H2 mutants kept *c.a.* 30% of their initial ABTS-activity values, and those temperatures were selected for the screening. At this point, it is important to highlight that both VP and HRPL libraries were functionally secreted by yeast, which made the development of the assay faster, reliable and simple since it was not necessary to include cell lysis steps, thus avoiding possible interferences with the complex lysate mixture. Besides, both VP and HRPL secretion levels were previously improved by several iterative rounds of directed evolution to provide: i) short incubations times for functional secretion



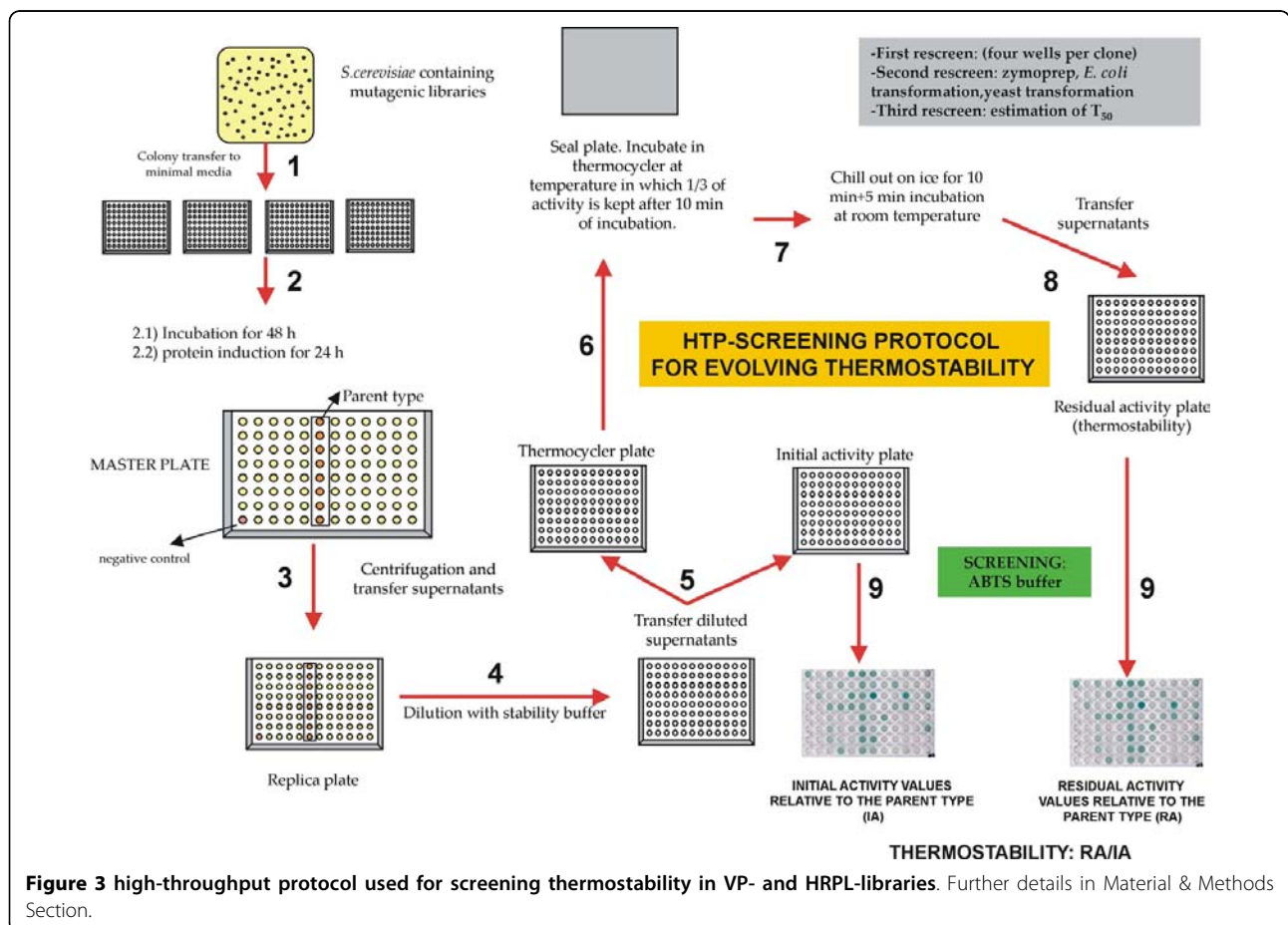
(24 h) after protein induction that becomes essential for the episomal plasmid stability; and ii) a high level of activity in supernatants (getting measurements from kinetic mode in a few seconds with turnover rates ranging from 0.5 to 1 ABTS-Units/mL). Taking advantage of the high level of activity in the culture broth, supernatants of respective libraries were previously diluted in suitable corresponding stability buffers to correctly assess initial and residual activities (Fig. 3). The final values of thermostability came from the ratio of residual activity to initial activity (RA/IA) normalized with the corresponding parent type. The initial activity value reflected the total activity of the mutant and was highly useful to prevent the selection of mutants with improved stability by greatly reduced activity.

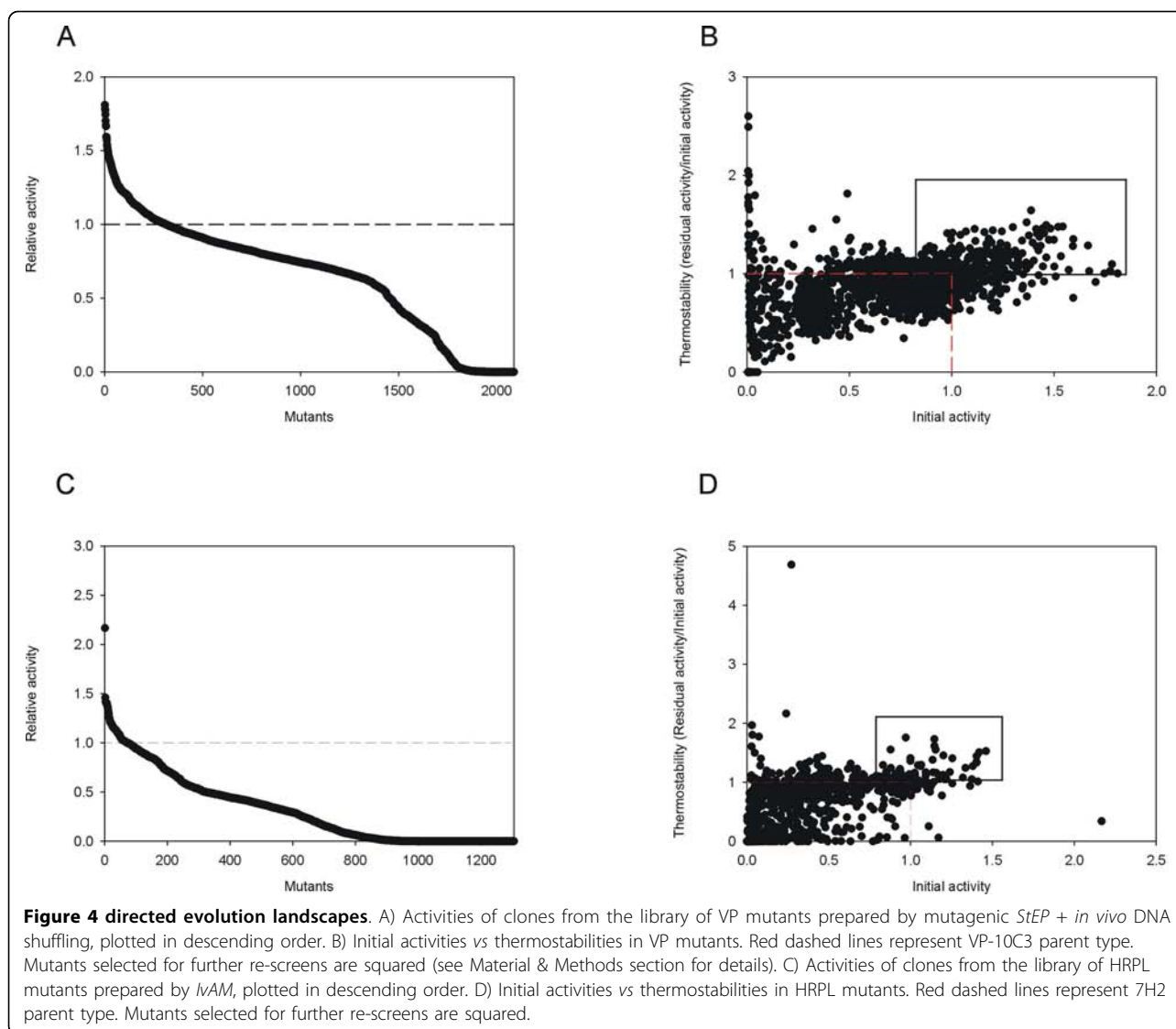
### 3. Library analysis

Under the above premises, two libraries of ~2000 clones were independently constructed and explored for VP and HRPL (Fig. 4). The estimated coefficients of variance for the landscapes of initial activity and residual activity were below 12%. Three consecutive re-screens were incorporated to ensure the selection of mutant *hits*

in protein function. Systematically, the best 50 clones retaining ~0.7 fold the activity of parent type and showing improvements in thermostability were selected and double rescreened from supernatants and new fresh transformants. Finally, best mutants were subjected to a third rescreen to estimate their  $T_{50}$ ; *i.e.* the transition midpoint of the inactivation curve of the protein as a function of temperature, which in our case was defined as the temperature at which the enzyme loses 50% of its activity following incubation for 10 min.

The best four mutants of VP library, with significant improvements in their activities and/or thermostabilities, were sequenced (Fig. 5). These variants shared the common feature of incorporating 4 mutations (E37K, V160A, T184M and Q202L) accumulated round after round of evolution in the same mature protein. Interestingly, the best variant of activity (R4 mutant with  $3180 \pm 30$  ABTS-Units/L) only contained the mentioned 4 mutations after 4 cycles of evolution. The remaining selected variants displayed a negative epistatic effect, *i.e.* the general combination of mutations is beneficial but at least one individual mutation is not, after incorporating new point mutations (with improvement ranging from 1.7 to 1.1



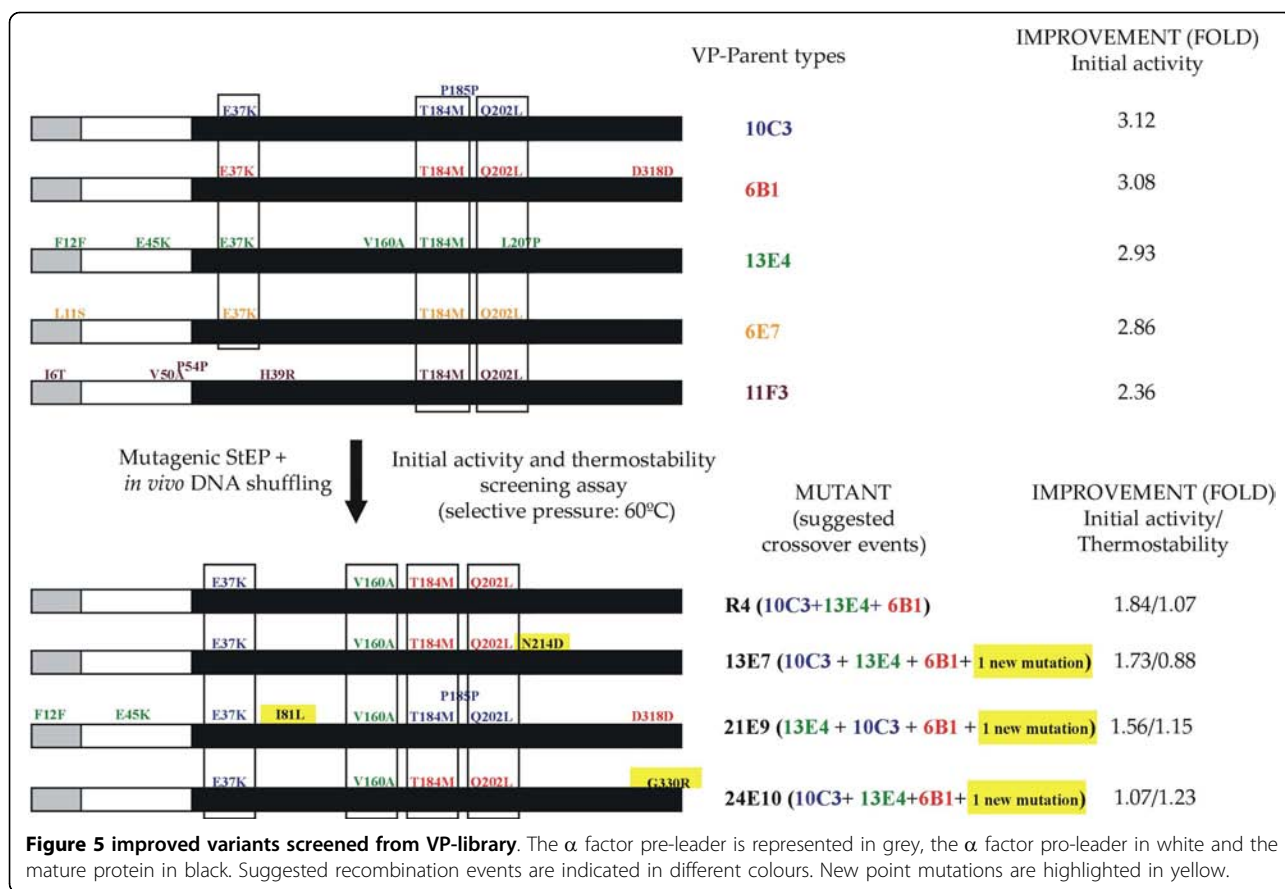


fold vs best parent type, 10C3 mutant). It is noteworthy that the best thermostable variant (24E10 mutant, with 1.23-fold better stability than 10C3) contains the same four mutations as R4 plus mutation G330R, which logically is responsible for the improvement in the thermostability. Mutation G330R shifted the  $T_{50}$  2.2°C, from 60.5°C to 62.7°C (Fig. 6A), but decreased the activity of the variant almost 0.77 fold. Indeed, the improvement in protein stability came at the cost of reducing activity (the well known tradeoff that usually appears between activity and stability for many single point mutations). However, the recombination method designed for this experiment (mutagenic *StEP* + *in vivo* DNA shuffling) made possible to join the four beneficial mutations for the activity which somehow buffered the effect of incorporating the stabilizing mutation G330R, giving rise to an enzyme with similar activity to the parent types but more

thermostable. Mutation G330R is placed at the C-terminal tail of VP, a controversial region that, although is not involved in VP oxidation of  $Mn^{2+}$  as initially suggested for the C-tail of *Phanerochaete chrysosporium* MnP [41], presents a high mobility that prevented to fix the position of the last 12 residues when the VP crystal structure was solved (PDB entry 2BOQ). This highly-mobile region could contribute to the thermal denaturation of the protein, and it is possible to speculate that the introduction of an arginine side-chain at position-330 could stabilize the region, and the whole VP protein, by establishing some new interaction/s, whose nature cannot be determined due to the lack of valuable crystallographic data for the C-terminal tail of VP.

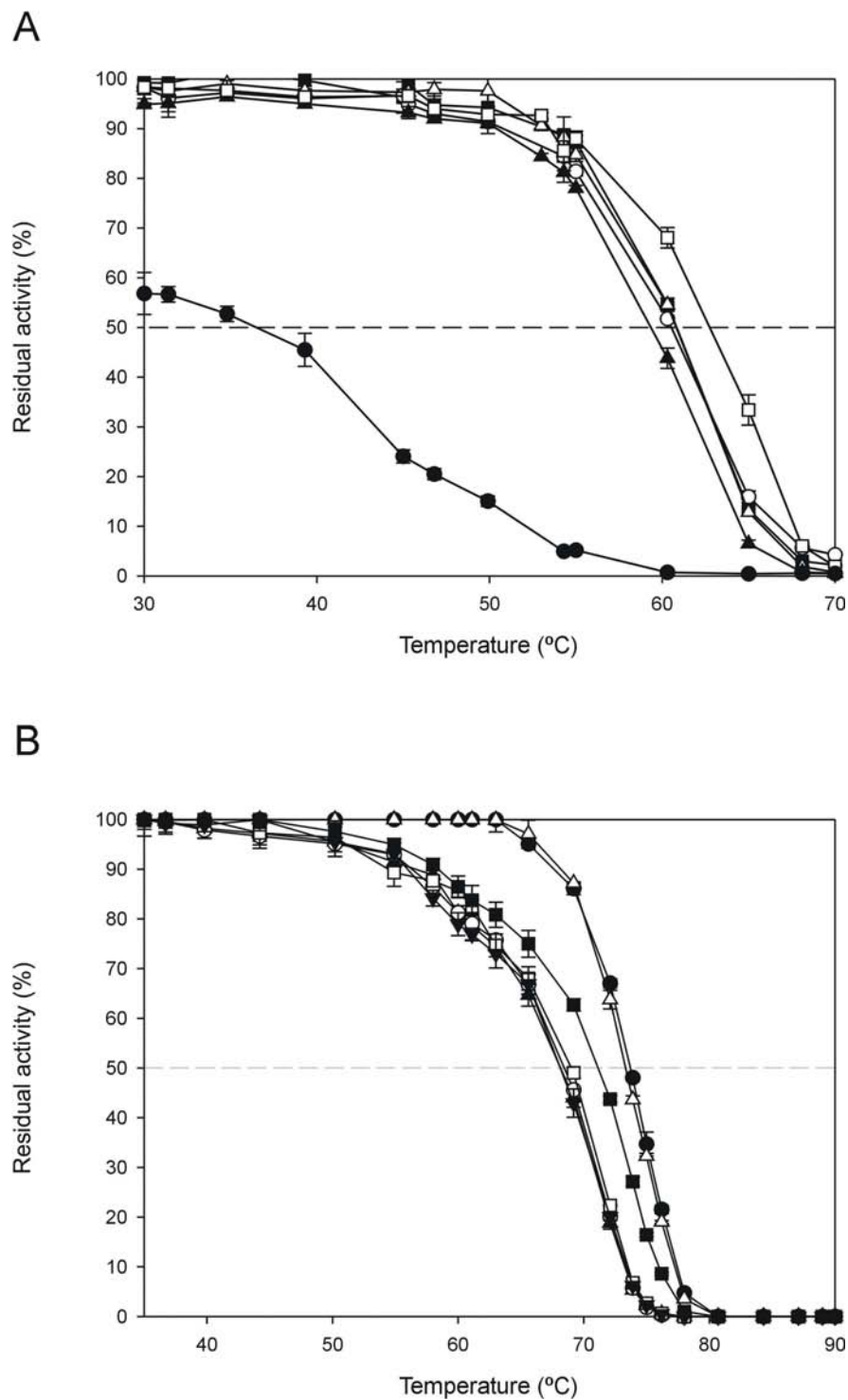
HRPL-library was generated by *IvAM* from 7H2 mutant, the best variant of the 5<sup>th</sup> round of evolution for total activity enhancement (Fig. 7). 7H2 was a good



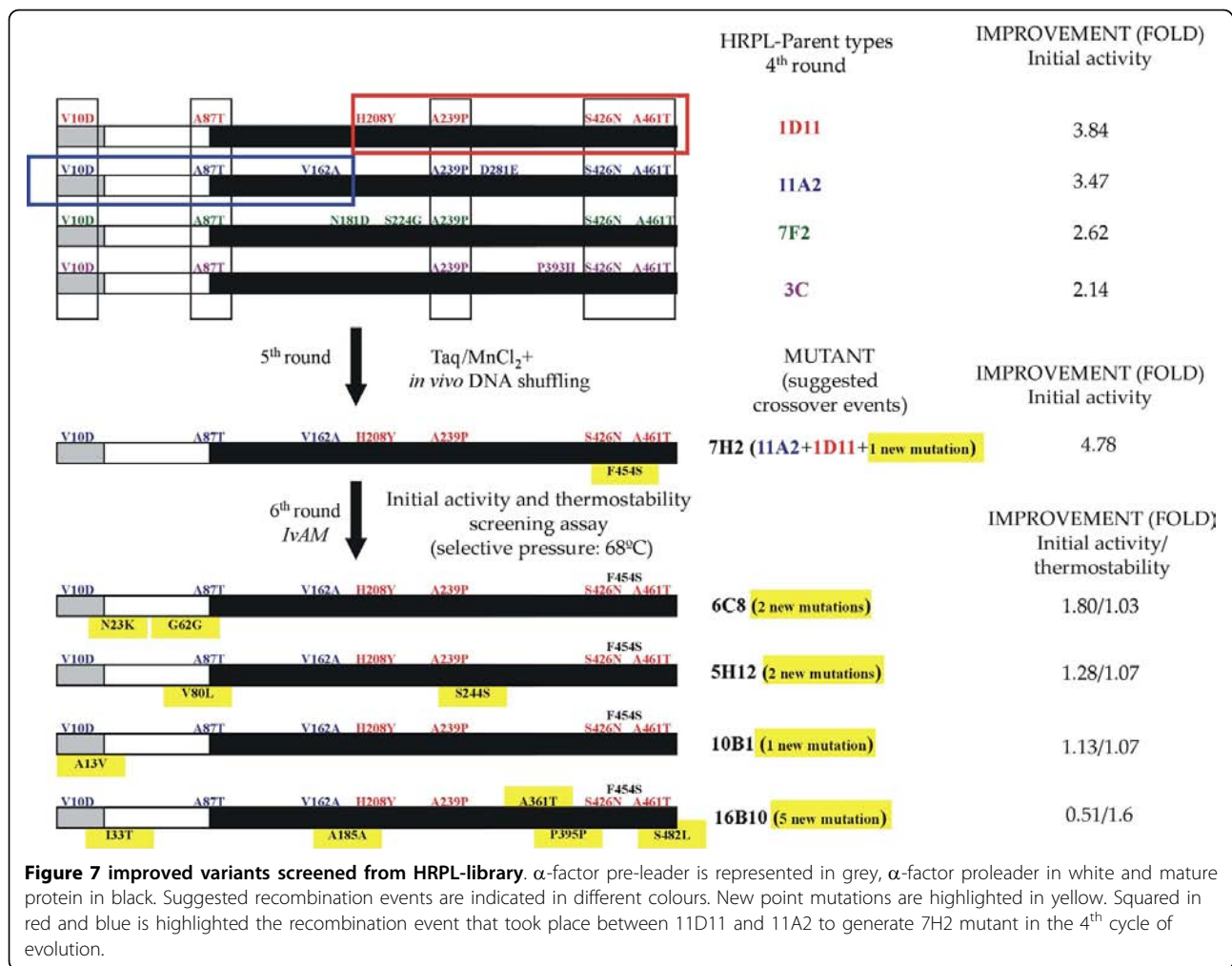


candidate to improve the thermostability: during the former round of evolution by mutagenic *in vivo* DNA-shuffling, although its activity was improved almost 5-fold (achieving  $1000 \pm 24$  ABTS-Units/L), its stability diminished considerably, with a decrease in the  $T_{50}$  of  $\sim 5^\circ\text{C}$  vs 1D11 and 11A2 parent types (Fig. 6B). As main consequence of the drop in the  $T_{50}$ , 7H2 was unstable during long-term storage (losing about 30% of its activity after 15 days at  $4^\circ\text{C}$ ). In the context of directed evolution experiments, this effect is not surprising and there are many examples in literature about falls in stability because of introducing beneficial but destabilizing mutations for enhanced activity [3]. Taking into account that 7H2 was created from a single crossover event between 1D11 and 11A2 variants (Fig. 7), and that both parent types shares similar  $T_{50}$  values ( $73.7$  and  $73.3^\circ\text{C}$  respectively, Fig. 6B) it became clear that the only new mutation incorporated in 7H2 (F454S) was responsible for the dramatic drop in the  $T_{50}$ . The mutation was mapped in a model based on the *Trametes trogii* laccase (97% identity) crystal structure [34]. Phe454 is placed in a region close to the T1 copper site, the place where the reducing substrate binds. The change from Phe to Ser at this position seems to disrupt several interactions with neighboring

residues, which probably affects stability. Best mutants of the thermostability cycle where sequenced (Table 1) and in all cases new mutations at the pre- or pro-leader were introduced benefiting the secretion levels by yeast (with improvement in total activities ranging from 1800 to 500 ABTS-Units/L). The best thermostable mutant (16B10 variant) displayed 1.6-fold better stability than 7H2, shifting its  $T_{50}$  value over  $3^\circ\text{C}$ , but at the cost of reducing its activity by half (Fig. 6B and Fig. 7). In spite of the fact that *IvAM* method takes advantage of mixing different mutational profiles created by polymerases with broad differences in mutational bias (Table 1), the constraint of starting from only one single parent type hindered to find appropriate crossover events (as happened in VP-libraries) that otherwise could have helped to prevent the detrimental effect on activity of introducing stabilizing mutations. 16B10 harbours 4 mutations in mature protein (two synonymous). Mutations A361T and S482L are placed at the surface of the protein, close to the C-end. Both mutations are at least  $21 \text{ \AA}$  from the nearest catalytic copper atom (Table 1, Fig. 8). Ala361 is located in a loop and interacts with Leu364 by a hydrogen bond. The change from Ala to Thr at position 361, kept the mentioned interaction and allowed one additional hydrogen



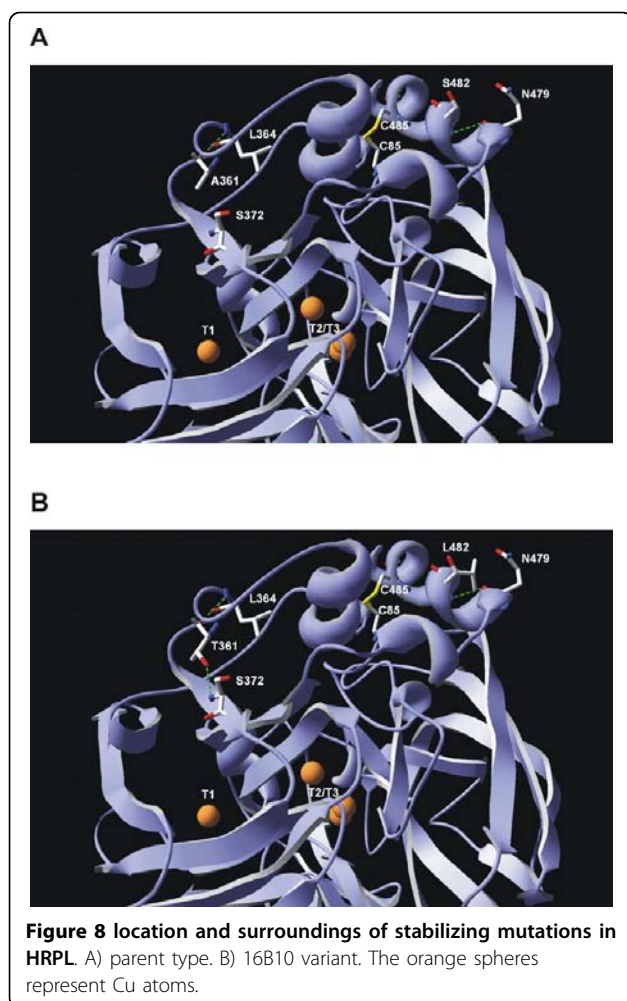
**Figure 6 A)  $T_{50}$  for VP parent types and different mutants of the evolutionary process.** White squares, 24E10 mutant; black triangles, R4 mutant; black squares, 10C3 mutant (best parent type from 3<sup>rd</sup> generation); white circles, best parent type from 2<sup>nd</sup> generation; white triangles, best parent type from 1<sup>st</sup> generation; black circles, native VP expressed in *E. coli* after *in vitro* refolded from inclusion bodies. B)  $T_{50}$  for HRPL parent types and mutants of the *in vitro* evolution. Black circle, 1D11 mutant (4<sup>th</sup> generation); white triangle, 11A2 mutant (4<sup>th</sup> generation); black triangle, 7H2 mutant (5<sup>th</sup> generation); white circle, 6C8 mutant (6<sup>th</sup> generation); white square, 5H12 mutant (6<sup>th</sup> generation); black down triangle, 10B1 mutant (6<sup>th</sup> generation); black square, 16B10 mutant (6<sup>th</sup> generation). Each point, including the standard deviation, comes from three independent experiments.



**Table 1 HRPL selected mutants generated by *IvAM***

Mutant	Amino acid Substitution	Nucleotide change	Mutation type	Location	Secondary structure motif	Distance to the T1 Site (Å)	Distance to the T2/T3 (Å)
6C8	<u>N23K</u>	AAC69AAA	Tv	Pro-leader			
	<u>G62G</u>	<b><u>6GGG186GGA</u></b> <sub>11</sub>	Ts	Pro-leader			
5H12	<u>V80L</u>	GTA238CTA	Tv	Pro-leader			
	<u>S244S</u>	<b><u>8TCG1005TCC</u></b> <sub>14</sub>	Tv	Mature protein			
10B1	<u>A13V</u>	GCA38GTA	Ts	Pre-leader			
	<u>I33T</u>	ATT98ACT	Ts	Pro-leader			
	<u>A185A</u>	<b><u>21GCT828GCC</u></b> <sub>13</sub>	Ts	Mature protein			
16B10	<u>A361T</u>	GCG1354ACG	Ts	Mature protein	loop	23	21
	<u>P395P</u>	<b><u>7CCC1458CCT</u></b> <sub>13</sub>	Ts	Mature protein			
	<u>S482L</u>	TCG1718ITG	Ts	Mature protein	sheet	34	24

Synonymous mutations are underlined; nucleotide change is highlighted in bold; subscript numbers on codons indicate codon usage. Tv, transversions; Ts, transitions.



bond with Ser372 placed in a beta-sheet. This structural reinforcement upon mutation might confer rigidity to the protein enhancing its stability. Mutation S482L is in the neighbourhood of the disulfide bridge between Cys485 and Cys85. Ser482 is part of a  $\alpha$ -helix and establishes a hydrogen bond with Gln479 of same motif. Inspection of the protein model suggests that the S482L mutation does not interrupt such a bond. The change of a polar amino acid by a bigger hydrophobic one might allow establishing hydrophobic interactions with surrounding residues which may further stabilize the protein structure at this region.

## Conclusions

In summary, *S. cerevisiae* is a valuable cell factory for the directed evolution of ligninolytic enzymes for thermostability and taking together, the VP and the HRPL evolved variants share several common features. First, the thermostability improvements obtained for both VP and HRPL systems are especially significant near the enzyme inactivation temperatures: the best VP (24E10) showed

~30% of its maximal activity at 65°C (3-fold more than the initial VP) and the best laccase (16B10) up to 40% of its maximal activity at 72°C (over 10-fold more than the corresponding parent type). Second, an apparently inherent tradeoff between activity and stability appeared in both enzymes for different amino acid substitutions. Although not physically incompatible, in general protein scaffolds activity and thermostability tend to act as communicating vessels and the laboratory design of any of them usually come at the cost of its counterpart. For protein engineers, to find single mutations which improve both properties simultaneously is extremely difficult. In nature, stability is under selection just in the case that it is required for biochemical function, hence mutations which join activity and stability are rare taking into account the genetic drift and that a selective pressure towards both features at the same time is not frequently exerted. It has been reported that in principle is easier to evolve thermostability while keeping activity than vice versa, although recent research indicates that evolving activity while maintaining stability can be accomplished as well [1,39]. We have demonstrated that the generation of complex crossover events along with the introduction of new mutations facilitates the improvement in the stability of ligninolytic oxidoreductases buffering the drops on their activities. In the evolutionary scenario, the recombination methods described in this work for the generation of diversity along with the screening assay engineered for this specific task can be valuable tools not only to tailor thermostable ligninolytic oxidoreductases but also other enzymatic systems.

## Abbreviations

ABTS: 2,2'-azino-bis (3 ethylbenzothiazoline 6 sulfonic acid); DMSO: dimethylsulfoxide; HRPL: High Redox Potential Laccases; IvAM: *In vivo* Assembly of Mutant libraries with different mutational spectra; RA/IA: Residual Activity/Initial Activity; StEP: Staggered Extension Process;  $T_{50}$ : Temperature at which the enzyme loses 50% of its activity following incubation for 10 minutes; VP: Versatile Peroxidase.

## Acknowledgements

Authors truly thank Prof. Ramón Santamaria from Salamanca University for providing PM1 laccase gene. This material is based upon work funded by National Projects CCG08-CSIC/PPQ-3706 and CSIC 200880I033; and EU Projects NMP4-SL-2009-229255 and NMP2-CT-2006-026456. D.M. thanks the CSIC for a JAE contract.

## Author details

<sup>1</sup>Department of Biocatalysis, Institute of Catalysis, CSIC, 28049 Madrid, Spain. <sup>2</sup>Centro de Investigaciones Biológicas, CSIC, 28040 Madrid, Spain.

## Authors' contributions

The PhD students EGR and DM carried out all the experiments on VP and HRPL respectively under the supervision of MA. ATM provided background on VP and co-supervised EGR. MA wrote the first draft which was revised by ATM and AB. MA coordinated the final version of the paper, which was read and approved by all authors.

## Competing interests

The authors declare that they have no competing interests.



Received: 3 December 2009 Accepted: 18 March 2010  
Published: 18 March 2010

## References

1. Bloom JD, Labthavikul ST, Otey CR, Arnold FH: **Protein stability promotes evolvability.** *Proc Natl Acad Sci USA* 2006, **103**:5869-5874.
2. Bloom JD, Wilke CO, Arnold FH, Adami C: **Stability and the evolvability of function in a model protein.** *Biophys J* 2004, **86**:2758-2764.
3. Bloom JD, Arnold FH: **In the light of directed evolution: Pathways of adaptive protein evolution.** *Proc Natl Acad Sci USA* 2009, **106**:9995-10000.
4. Watanabe K, Ohkuri T, Yokobori S, Yamagishi A: **Designing thermostable proteins: Ancestral mutants of 3-isopropylmalate dehydrogenase designed by using a phylogenetic tree.** *J Mol Biol* 2006, **355**:664-674.
5. Ogino H, Ishikawa H: **enzymes which are stable in the presence of organic solvents.** *J Biosci Bioeng* 2001, **91**:109-116.
6. Arnold FH: **Engineering enzymes for nonaqueous solvents.** *Trends Biotechnol* 1990, **8**:244-249.
7. Suen WC, Zhang NY, Xiao L, Madison V, Zaks A: **Improved activity and thermostability of *Candida antarctica* lipase B by DNA family shuffling.** *Protein Eng* 2004, **17**:133-140.
8. Johannes TW, Woodyer RD, Zhao HM: **Directed evolution of a thermostable phosphite dehydrogenase for NAD(P)H regeneration.** *Appl Environ Microb* 2005, **71**:5728-5734.
9. Arnold FH, Georgiou G: **Directed Enzyme Evolution: screening and selection methods.** *Methods in Molecular Biology* Humana Press, Totowa, New Jersey 2003, **231**.
10. Reetz MT, Carballeira JD, Vogel A: **Iterative saturation mutagenesis on the basis of B factors as a strategy for increasing protein thermostability.** *Angew Chem Int Ed* 2006, **45**:7745-7751.
11. Bommarius AS, Broering JM, Chaparro-Riggers JF, Polizzi KM: **High-throughput screening for enhanced protein stability.** *Curr Opin Biotech* 2006, **17**:606-610.
12. López Camacho C, Salgado J, Lequerica JL, Madarro A, Ballestar E, Franco L, Polaina J: **Amino acid substitution enhancing thermostability of *Bacillus polymixa* beta-glucosidase A.** *Biochem J* 1996, **314**:833-838.
13. Miyazaki K, Takenouchi M, Kondo H, Noro N, Suzuki M, Tsuda S: **Thermal stabilization of *Bacillus subtilis* family-11 xylanase by directed evolution.** *J Biol Chem* 2006, **281**:10236-10242.
14. Giver L, Gershenson A, Freskgard PO, Arnold FH: **Directed evolution of a thermostable esterase.** *Proc Natl Acad Sci USA* 1998, **95**:12809-12813.
15. Arnold FH, Giver L, Gershenson A, Zhao HM, Miyazaki K: **Directed evolution of mesophilic enzymes into their thermophilic counterparts.** *Proc Natl Acad Sci USA* 1999, **870**:400-403.
16. Hao J, Berry A: **A thermostable variant of fructose bisphosphate aldolase constructed by directed evolution also shows increased stability in organic solvents.** *Protein Eng* 2004, **17**:689-697.
17. Morawski B, Quan S, Arnold FH: **Functional expression and stabilization of horseradish peroxidase by directed evolution in *Saccharomyces cerevisiae*.** *Biotechnol Bioeng* 2001, **76**:99-107.
18. Salazar O, Cirino PC, Arnold FH: **Thermostabilization of a cytochrome P450 peroxygenase.** *Chem Bio Chem* 2003, **4**:891-893.
19. Reading NS, Aust SD: **Engineering a disulfide bond in recombinant manganese peroxidase results in increased thermostability.** *Biotechnol Progr* 2000, **16**:326-333.
20. Sun LH, Petrounia IP, Yagasaki M, Bandara G, Arnold FH: **Expression and stabilization of galactose oxidase in *Escherichia coli* by directed evolution.** *Protein Eng* 2001, **14**:699-704.
21. Kunamneni A, Camarero S, García-Burgos C, Plou FJ, Ballesteros A, Alcalde M: **Engineering and Applications of fungal laccases for organic synthesis.** *Microb Cell Fact* 2008, **7**.
22. Alcalde M, Ferrer M, Plou FJ, Ballesteros A: **Environmental biocatalysis: from remediation with enzymes to novel green processes.** *Trends Biotechnol* 2006, **24**:281-287.
23. Martínez AT, Ruiz-Dueñas FJ, Martínez MJ, del Río JC, Gutiérrez A: **Enzymatic delignification of plant cell wall: from nature to mill.** *Curr Opin Biotechnol* 2009, **20**:348-357.
24. Alcalde M: **Laccase: biological functions, molecular structure and industrial applications.** *Industrial Enzymes: structure, functions and applications* New York, Springer Polaina J, MacCabe AP 2007, 459-474.
25. Riva S: **Laccases: blue enzymes for green chemistry.** *Trends Biotechnol* 2006, **24**:219-226.
26. Ruiz-Dueñas FJ, Morales M, García E, Miki Y, Martínez MJ, Martínez AT: **Substrate oxidation sites in versatile peroxidase and other basidiomycete peroxidases.** *J Exp Bot* 2009, **60**:441-452.
27. Ruiz-Duenas FJ, Martinez MJ, Martinez AT: **Heterologous expression of *Pleurotus eryngii* peroxidase confirms its ability to oxidize Mn<sup>2+</sup> and different aromatic substrates.** *Appl Environ Microbiol* 1999, **65**:4705-4707.
28. Coll PM, Taberner C, Santamaria R, Perez P: **Characterization and structural analysis of the laccase I gene from the newly isolated ligninolytic basidiomycete PM1 (CECT 2971).** *Appl Environ Microbiol* 1993, **59**:4129-4135.
29. García E, Martínez MJ, Dueñas J, Martínez AT, Alcalde M: **High redox potential peroxidases engineered by directed evolution.** *Patent n° 200930157.* Spain 2009.
30. Zhao HM, Giver L, Shao ZX, Affholter JA, Arnold FH: **Molecular evolution by staggered extension process (StEP) *in vitro* recombination.** *Nature Biotech* 1998, **16**:258-261.
31. Okkels JS: ***In vivo* gene shuffling in yeast: a fast and easy method for directed evolution of enzymes.** *Enzyme Functionality: Design, Engineering and Screening* Marcel Dekker, Inc. New York Svendsen A 2004, 413-424.
32. Zumárraga M, Camarero S, Shleev S, Martínez-Arias A, Ballesteros A, Plou FJ, Alcalde M: **Altering the laccase functionality by *in vivo* assembly of mutant libraries with different mutational spectra.** *Proteins* 2008, **71**:250-260.
33. Zumárraga M, Domínguez CV, Camarero S, Shleev S, Polaina J, Martínez-Arias A, Ferrer ML, de Lacey A, Fernández V, Ballesteros A, Plou FJ, Alcalde M: **Combinatorial Saturation Mutagenesis of the Myceliophthora thermophila laccase T2 mutant: the connection between the C-terminal plug and the conserved 509VSG511 tripeptide.** *Comb Chem High T Scr* 2008, **11**:807-816.
34. Matera I, Gullotto A, Tili S, Ferraroni M, Scozzafava A, Briganti F: **Crystal structure of the blue multicopper oxidase from the white-rot fungus *Trametes trogii* complexed with *p*-toluate.** *Inorg Chim Acta* 2008, **361**:4129-4137.
35. Shuster JR: **Gene expression in yeast: protein secretion.** *Curr Opin Biotechnol* 1991, **2**:685-690.
36. Tracewell CA, Arnold FH: **Directed enzyme evolution: climbing fitness peaks one amino acid at a time.** *Curr Opin Chem Biol* 2009, **13**:3-9.
37. Cherry JR, Lamsa MH, Schneider P, Vind J, Svendsen A, Jones A, Pedersen AH: **Directed Evolution of a fungal peroxidase.** *Nature Biotech* 1999, **17**:379-384.
38. Bulter T, Alcalde M, Sieber V, Meinhold P, Schlachtbauer C, Arnold FH: **Functional expression of a fungal laccase in *Saccharomyces cerevisiae* by directed evolution.** *Appl Environ Microb* 2003, **69**:987-995.
39. Zumárraga M, Bulter T, Shleev S, Polaina J, Martínez-Arias A, Plou FJ, Ballesteros A, Alcalde M: ***In vitro* evolution of a fungal laccase in high concentrations of organic cosolvents.** *Chem Biol* 2007, **14**:1052-1064.
40. Alcalde M, Zumárraga M, Polaina J, Ballesteros A, Plou FJ: **Combinatorial saturation mutagenesis by *in vivo* overlap extension for the engineering of fungal laccases.** *Comb Chem High T Scr* 2006, **9**:719-727.
41. Ruiz-Dueñas FJ, Morales M, Pérez-Boada M, Choinowski T, Martínez MJ, Piontek K, Martínez AT: **Manganese oxidation site in *Pleurotus eryngii* versatile peroxidase: A site-directed mutagenesis, kinetic, and crystallographic study.** *Biochemistry* 2007, **46**:66-77.

doi:10.1186/1475-2859-9-17

Cite this article as: García-Ruiz et al.: Evolving thermostability in mutant libraries of ligninolytic oxidoreductases expressed in yeast. *Microbial Cell Factories* 2010 **9**:17.